

Effect of Potassium Nitrate and Salinity on Growth and Endogenous Gibberellins of *Glycine max* Var. Daewonkong.

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ABSTRACT:

Plant growth and development was affected greatly through salinity. Appropriate nutrients application on salinity condition can reduce its harmful effects and increased productivity of salinized soils. A pot experiment was carried out in 2007 in cs Lab., Department of Agronomy, Kyungpook National University, Korea to assess the effects of potassium nitrate on *Glycine max* (Var. Daewonkong) under different salinity levels. Experiment was complete randomized block design (CRBD), consisted of 9 treatments with 3 replications per treatment and each replication comprising 9 plants. KNO₃ and NaCl were applied to soybean plants 14 days after sowing (14 DAS). KNO₃ was applied at the rates of 5.0 mM and 10.0 mM, each pot receiving 150 ml of KNO₃ solution. For salt stress induction, single dose of 300 ml of NaCl solution of 70 mM and 140 mM strength was given to the plants. Plant length, shoot and root fresh and dry weights were measured for harvested soybean plants while chlorophyll content was estimated in fully expanded leaves. Plant samples were harvested after 24 h of KNO₃ and NaCl application for the analysis of endogenous bioactive Gibberellin. Results showed that all growth parameters (shoot length, fresh and dry shoot weight, fresh and dry root weight), chlorophyll content and endogenous bioactive GA₁ and GA₄ content of soybean cultivar Daewonkong were reduced under NaCl stress. KNO₃ application enhanced shoot length, fresh and dry shoot weight, fresh and dry root weight, chlorophyll contents and endogenous bioactive GA₁ and GA₄ content as compared to control treatment.

KEYWORDS: Salinity, Potassium nitrate, Endogenous Gibberellin, Chlorophyll.

INTRODUCTION:

Salinity is a serious problem that limiting the productivity of crop plants. Crop production rate decreases due to salinity toxicity every year [1]. It results from high level of soluble salts in the soil. High amounts of dissolved salts in soils can limit crop yield [2]. High concentration of salts causes hyper osmotic stress and ion imbalance that produce secondary effects on plants growth [3] [4]. Salt-affected soils currently account for 8% of the world's total land area [5], and the salt-affected agricultural land is predicted to double by 2050 for irrigated agriculture and some semi-arid areas [6] [7]. Most of the salt stresses in nature are due to Na^+ salts, particularly NaCl [8]. High salinity lowers water potential and induces ionic stress, and results in secondary oxidative stress. It severely limits growth and development of plants by affecting different metabolic processes such as CO_2 assimilation, oil and protein synthesis [9].

One approach to minimize effects of salinity is use of nutrient application or nutrient enrichment into soil or soilless culture to increase tolerance to plant salinity by alleviating Na^+ and Cl^- injury to plants [10]

[11] [12]. Potassium (K) acts as a very essential and important nutrient for the plant growth and development. It is necessary in plants to improve the efficiency of photosynthesis and use of water [13]. In wheat, deficiency of K^+ causes so many problems and shows many deficiency symptoms in plants growth such as weaker straw, increased lodging and decrease in growth [13]. Application of K to wheat plants under saline conditions enhances the growth of the plants and decreases the effect of salinity [14].

Soybean (*Glycine max*) is a major agricultural crop that is used widely for providing human and animal food because of its high oil and protein content (18 and 38%, respectively) [15]. It is classified as a moderately salt-sensitive crop [2]. Its seed is a major source of high-quality protein and oil for human consumption [16]. Its chemical composition has made it one of the most valuable agronomic crops worldwide [17]. Its protein has great potential as a major source of dietary protein. The oil produced from soybean is highly digestible and contains no cholesterol [18]. Growth, development and yield of soybean are the result of genetic potential interacting with environment.

Objectives:

This experiment was carried out in order to assess the effectiveness of combined supplementary K and N (as KNO_3) for overcoming salinity stress. The specific aim was to determine if potassium nitrate (KNO_3) would correct K and N deficiencies in the presence of high NaCl, and also to assess effects of combined K and N on soybean growth, chlorophyll content and endogenous gibberellins.

MATERIALS AND METHODS:

This experiment was complete randomized block design (CRBD), consisted of 9 treatments with 3 replications per treatment and each replication comprising 9 plants. Seeds of high yielding Korean soybean cultivar Daewonkong were procured from Plant Genetics Lab., Department of Agronomy, Kyungpook National University, Korea.

General Procedure:

Seeds of soybean cultivar Daewonkong were surface sterilized with 5% NaClO for 15 min and then thoroughly rinsed with double de-ionized distilled water. Seeds

were sown in plastic pots (5.5 liter) filled with horticulture soil on 25th June, 2007. The composition of horticulture soil was as follows; peat moss (13-18%), perlite (7-11%), coco-peat (63-68%) and zeolite (6-8%), while the macro-nutrients were present as follows: NH_4^+ ~90 mg/L; NO_3^- ~205 mg/L; P_2O_5 ~350 mg/L and K_2O ~100 mg/L. Initially 5 seeds were sown, that were later thinned to three seedlings per pot. The experiment was conducted under controlled greenhouse environment. Soybean plants were harvested on 25th July, 2007 (30 DAS).

Concentration and Time of KNO_3 and NaCl

Application:

KNO_3 and NaCl were applied to soybean plants 14 days after sowing (14DAS). KNO_3 was applied at the rates of 5.0 mM and 10.0 mM, each pot receiving 150 ml of KNO_3 solution. For salt stress induction, single dose of 300 ml of NaCl solution of 70 mM and 140 mM strength was given to the plants.

Analysis of Endogenous Bioactive

Gibberellin:

Plant samples were harvested after 24 h of KNO_3 and NaCl application and were

immediately frozen in liquid nitrogen and stored at minus 80°C. Plant samples were lyophilized in freeze drier (Virtis, SP Industries Inc.). Leaves of the lyophilized plant samples were crushed to powder form and used for analysis of endogenous bioactive gibberellin.

Extraction and quantification methods:

Measurement of Growth Component:

Plant length, shoot and root fresh and dry weights were measured for harvested soybean plants while chlorophyll content of fully expanded leaves was analyzed with the help of chlorophyll meter (Minolta Co., Ltd, Japan). Four replicates of 6 plants each per treatment were randomly selected for measuring growth parameters. Dry weights were measured after drying the samples at 70°C for 48 h in an oven.

Statistical Analysis of Data:

The data was subjected to analysis of variance (ANOVA SAS release 9.1; SAS, NC, USA) and Duncan's multiple range test (DMRT).

RESULTS AND DISCUSSION:

Effect of KNO₃ on Soybean Growth under Salt Stress:

Many factors involved in lowering the crop growth and yield, such as drought, salinity, and high temperatures [19] [20]. Salinity adversely affect the plant growth and development [21]. Our study showed that all growth parameters (shoot length, fresh and dry shoot weight, fresh and dry root weight) of soybean cultivar Daewonkong were reduced under NaCl stress (Table 1). Decrease in plant lengths was done by the effect of salinity and this phenomenon was observed by a number of investigators [22] [23] [24]. Similar investigations were also reported by kausar et al., [25]. They find out that lengths of plants were adversely affected by salinity stress. Growth of wheat plant was reduced by lower fixation of carbon dioxide, reduced stomatal conductance, photochemical and biochemical capacity or combinations of all other factors in stress environments [26]. Several reports are available in agreement of the present studies that fresh biomass production reduced by the salinity stress as compared to non-saline conditions [25] [27] [28]. Reduction in dry weight reflects the increased metabolic energy cost and reduced carbon gain. It also reflects salt impacts on tissues [29] reduction in

photosynthetic rates [30] [31] and attainment of maximum salt concentration tolerated by the fully expanded leaves [32].

KNO₃ application enhanced shoot length, fresh and dry shoot weight, fresh and dry root weight as compared to control treatment so its application alleviated the detrimental effect of NaCl on soybean growth parameters (Table 1). Foliar supply of KNO₃ to the salt treated plants may reduce toxic ions uptake as well improve K and N status of salt treated plants. Potassium has role in ionic balance which is reflected in nitrate metabolism [33]. Nitrogen plays role of chlorophyll and protein so act as an essential element for plant growth. Potassium application result an increase in leaf potassium content which was accompanied by increased rates of photosynthesis, photorespiration and RuBP carboxylase activity. Ebert *et al.*, [34] found that supplying of Ca(NO₃)₂ at 10 mM had a beneficial effect on growth and metabolism of NaCl treated guava seedlings. Akram *et al.*, [35] observed an improvement in growth of sunflower due to the foliar spray of K₂SO₄ and KNO₃ at 1.25% under saline concentration of 150 mM NaCl.

Present investigation confirmed that salt stress markedly decreases leaf chlorophyll

content during vegetative growth stage (Table 1). These results are in agreement with those of Azooz *et al.*, [36] for sorghum and Dager *et al.*, [37] for *Salvadora persica*. The reduction in leaf chlorophyll content under NaCl stress has been attributed to the destruction of chlorophyll pigments and the instability of the pigment protein complex [38]. Another cause for the reduction is the interference of salt ions with the de novo synthesis of proteins, the structural component of chlorophyll, rather than the breakdown of chlorophyll [39]. Potassium and nitrogen supplemented in the form of KNO₃ can significantly improve the variables affected by high salinity and can also correct both K and N deficiencies. Addition of KNO₃ successfully ameliorated leaf chlorophyll and carotenoid contents of plant grown in salt stress. Stress conditions result in limited photosynthesis due to a decline in Rubisco activity and reduced gas exchange [40]. Application of SA or KNO₃ to salt and water stress treated barley plants enhanced the contents of photosynthetic pigments. The decrease Na⁺ and increase K⁺ in stressed barley plants in response to application of SA or KNO₃ may be caused maintenance of photosynthesis.

Table 1. Effect of KNO₃ and salinity on growth parameters and chlorophyll content of *Glycine max* Var. Daewonkong

Treatment	Solution strength (mM)	Plant length (cm)		Shoot weight (g plant ⁻¹)		Root weight (g plant ⁻¹)		Chl. content (plant ⁻¹)
		Shoot	Root	F.W	D.W	F.W	D.W	
Control	0	38.2±0.7 ^a	44.9±1.2 ^a	7.19±0.4 ^{ab}	1.16±0.04 ^{abc}	2.22±0.2 ^{ab}	0.28±0.03 ^a	28.3±0.5 ^{ab}
NaCl	70	37.3±1.0 ^{ab}	44.8±7.5 ^a	6.38±0.6 ^{bc}	0.75±0.1 ^{cd}	2.19±0.02 ^{ab}	0.24±0.02 ^a	25.4±1.1 ^{abcd}
	140	34.6±1.5 ^b	40.2±2.3 ^a	4.05±0.9 ^c	0.64±0.14 ^d	1.73±0.3 ^b	0.21±0.01 ^a	22.0±2 ^{cd}
KNO ₃	5.0	39.5±1.4 ^{ab}	37.3±6.5 ^a	7.84±0.6 ^{ab}	1.19±0.09 ^{abc}	2.76±0.6 ^{ab}	0.39±0.09 ^a	28.6±0.6 ^{ab}
	10.0	42.3±4.0 ^{ab}	45.1±5.9 ^a	9.3±1.7 ^a	1.36±0.2 ^{ab}	3.33±0.9 ^a	0.46±0.09 ^a	29.3±0.3 ^a
NaCl + KNO ₃	70+5.0	38.9±1.7 ^{ab}	44.7±3.8 ^a	9.07±0.2 ^a	1.4±0.01 ^a	3.12±0.2 ^{ab}	0.38±0.03 ^a	26.5±1 ^{abcd}
	70+10.0	40.3±0.9 ^{ab}	38.7±8.7 ^a	7.67±0.4 ^{ab}	1.0±0.08 ^{abcd}	2.29±0.02 ^{ab}	0.29±0.02 ^a	26.9±0.2 ^{abc}
KNO ₃	140+5.0	37.8±1.3 ^{ab}	43.1±7.0 ^a	6.25±0.3 ^{bc}	0.83±0.1 ^{cd}	2.23±0.2 ^{ab}	0.23±0.04 ^a	25.7±1.7 ^{abc}
	140+10.0	38.6±1.1 ^{ab}	43.7±5.9 ^a	6.79±0.3 ^{bc}	0.96±0.04 ^{cd}	1.87±0.1 ^{ab}	0.26±0.02 ^a	26.1±1.1 ^{bcd}

*In a column, treatment means having a common letter(s) are not significantly different at the 5% level by DMRT.

Endogenous Bioactive Gibberellin Content of Daewonkong:

The endogenous bioactive GA₁ and GA₄ content of soybean leaves showed a steady decline in response to elevated NaCl nutrition. Single application of KNO₃ greatly enhanced bioactive GA₁ and GA₄. Reduction of GA levels and signalling has been shown to contribute to plant growth restriction on exposure to several stresses, including cold, salt and osmotic stress [41]. The endogenous bioactive GA₁ and GA₄ content of soybean leaves showed a steady decline in response to elevated NaCl

nutrition. Single application of KNO₃ greatly enhanced bioactive GA₁ and GA₄. Application of KNO₃ to salt stressed plants yielded high GA₄ content while GA₁ content was not significantly affected; soybean thus showing a differential response in bioactive gibberellin metabolism to KNO₃ nutrition. Our results also demonstrated that application of KNO₃ was more effective at lower salt stress level as GA₁ and GA₄ contents in soybean leaves were higher with basic NaCl (70 mM) as compared to double NaCl (140 mM) treated plants. The concentration of GA₄ was much higher than GA₁ in all treatments (Fig. 1).

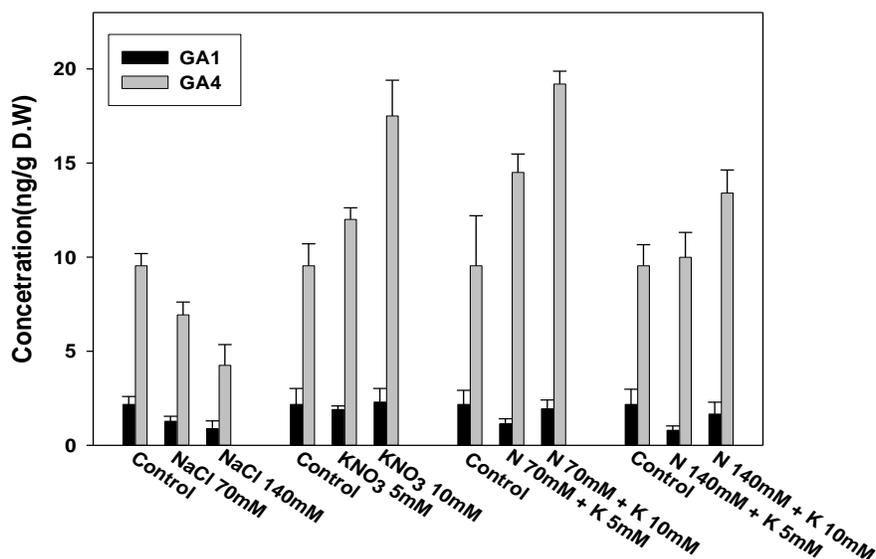


Figure 1 Level of endogenous bioactive GA₁ and GA₄ content in response to elevated KNO₃ and NaCl.

REFERENCES:

- [1] Hirt H and K Shinozaki. 2004. Plant Responses to Abiotic Stress. Springer, Berlin – Heidelberg - New York – Hong Kong – London – Milan – Paris – Tokyo
- [2] Munns R and M Tester. (2008) Mechanisms of salinity tolerance. *Annu. Rev. Plant Biol.* 59, 651–681.
- [3] Hasegawa PM, RA Bressan, JK Zhu and PBohernet. 2000. Plant cellular and molecular responses to high salinity. *Annu. Rev. Plant Physiol, Plant Mol. Boil.*, 51, 463-499.

- [4] Zhu JK. 2001. Plant salt tolerance. *Trends in plant Sci.* 61, 66-77. *Sci., Tech. and Dev.*, 32 (2): 99-103, 2013.
- [5] FAO A. 2000. Extent and causes of salt affected soils in participating countries. Available from <http://www.fao.org/ag/agl/agll/spush/topic2.htm>.
- [6] Pitman MG and A Läuchli. 2002. Global Impact of Salinity and Agricultural Ecosystems. In: *Salinity: Environment \x96 Plants \x96 Molecules* (Läuchli, A., Lüttge, U., eds). Dordrecht, the Netherlands: Kluwer, pp. 3–20.

- [7] Rengasamy P. 2006. World salinization with emphasis on Australia. J. Exp. Bot. 57, 1017–1023.
- [8] Demiral MA. 2005. Comparative response of two olive (*Olea europaea*) cultivars to salinity. Turk J. Agric. 25: 267-274.
- [9] Nasir Khan M, MH Siddiqui, F Mohammad, M Masroor, A Khan and M Naeem. 2007. Salinity induced changes in growth, enzyme activities, photosynthesis, proline accumulation and yield in linseed genotypes. World J. Agric. Sci., 3: 685- 695.
- [10] Alpaslan M, A Inal, SA Güne., V Çikili and H Özcan. 1999. Effect of zinc treatment on the alleviation of sodium and chloride injury in tomato (*Lycopersicon esculentum* [L.] Mill. cv. Lale) grown under salinity. Turkish Journal of Botany, 23: 1–6.
- [11] Pardossi A, G Bagnoli, F Malorgio, CA Campiotti and F Tognoni. 1999. NaCl effects on celery (*Apium graveolens* L.) grown in NFT. Scientia Horticulturae, 81: 229–242.
- [12] Tzortzakis NG. 2009. Influence of NaCl and calcium foliar spray on lettuce and endive growth using nutrient film technique. International Journal of Vegetable Science, 15: 1–13.
- [13] Ross MK. 2001. Potassium as fertilizer for plants. J. Plant Nutr. 425-433.
- [14] Safaa R, L El, T Magdi, H Abde and R Fatma. 2013. Effect of Potassium Application on Wheat (*Triticum aestivum* L.) Cultivars grown under salinity stress. World Applied Sciences Journal, 26 (7): 840-850, 2013.
- [15] Singh G. 2010. The Soybean: Botany, Production and Uses. Oxford shire, UK: CABI Publishing.
- [16] Katerji N, JW Van Hoorn, A Hamdy, M Mastrorilli, T Oweis and W Erskine. 2001. Response of two varieties of lentil to soil salinity. Agric. Water Management. 47: 179-190.
- [17] Thomas JMG, KJ Boote, LH Allen, M Gallo-Meagher and JM Davis. 2003. Seed physiology and metabolism: Elevated temperature and carbon dioxide effects on soybean seed composition and transcript abundance. Crop Sci., 43: 1548- 1557.
- [18] Essa TA and DH Al-ani. 2001. Effect of salt stress on the performance of six soybean genotypes. Pakistan Journal of Biological Sciences, 4: 175-177.
- [19] Bray EA, JB Serres and K Weretilny. 2000. Responses to abiotic stress. In B. Buchanan, W. Gruissem & R. Jones (Eds.), Biochemistry and molecular biology of plants. Plant Physiology, 1158-1203.
- [20] Wang M, Q Zheng, Q Shen, and S Guo. 2013. The critical role of potassium in plant

stress response. International Journal of Molecular Sciences, 14, 7370-7390.

[21] Gupta B, and B Haung. 2014. Mechanism of salinity tolerance in plants: Physiological, biochemical, and molecular characterization. International Journal of Genomics Article ID 701596. <http://dx.doi.org/10.1155/2014/701596>

[22] Khan AM, E Islam, MU Shirazi, M. U., Mumtaz, S., & Mujtaba, S. M. (2010). Physiological response of various wheat genotypes to salinity. Pakistan Journal of Botany, 42, 3497-3505.

[23] Ashraf M, M Afzal, R Ahmad, and S Ali. 2011. Growth and yield components of wheat genotypes as influenced by potassium and farm yard manure on a saline sodic soil. *Soil and Environment*, 30, 115-121.

[24] Ali A, SMA Basra, S Hussain, and J Iqbal. 2012. Increased Growth and Changes in Wheat Mineral Composition through Calcium Silicate Fertilization under Normal and Saline Field Conditions. *Chilean J. Agric. Res.*, 72(1).

[25] Kausar A, MY Ashraf, I Ali, M Niaz, and Q Abbass. 2012. Evaluation of sorghum varieties/lines for salt tolerance using physiological indices as screening tool. *Pakistan Journal of Botany*, 44(1), 47-52.

[26] Khan MH and SK Panda. 2008. Alterations in root lipid peroxidation and antioxidant responses in two rice cultivars under salinity stress. *Acta Physiology Plant*, 30, 89-91.

[27] Akhtar J, R Ahmad, MY Ashraf, A Tanveer, EA Waraich, and H Oraby. 2013. Influence of exogenous application of salicylic acid on salt stressed mung bean (*Vigna radiata*): growth and nitrogen metabolism. *Pakistan Journal of Botany*, 45, 119-125.

[28] Kausar A and M Gull. 2014. Nutrients uptake and growth analysis of four sorghum (*Sorghum bicolor* L.) genotypes exposed to salt stress. *Pensee Journal*, 76(4).

[29] Karimi J, M Ghorbanli, H Heidari, RA KhavariNejad and M Hassareh. 2005. The effects of NaCl on growth, water relations, osmolytes and ion content in *Kochiaprostrata*. *Biol. Planta*, 49: 301- 304.

[30] Ziska LH, JR Seemann and TM Dejong. 1990. Salinity induced limitation on photosynthesis in *Prunus salicina*, a deciduous tree species. *Plant Physiol.*, 93: 864- 870.

[31] Ashraf M. 2004. Some important physiological selection criteria for salt tolerance in plants. *Flora*, 199: 361-376.

- [32] Hu Y, HSchnyder and USchmidhalter. 2000. Carbohydrate deposition and partitioning in elongation leaves of wheat under saline soil conditions. *Aust. J. Plant Physiol.*, 27: 363- 370.
- [33] Jeschke WD and O Wolf.1985. Na dependent net K retranslocation in leaves of *Hordeum vulgare* cv. 'California Mariout' and *Hordeum distichon* cv. 'Villa' under salt stress. *J Plant Physiol* 121:211-223.
- [34] Ebert G, J Eberle, H Ali-Dinar and P Lüdders. 2002. Ameliorating effects of $\text{Ca}(\text{NO}_3)_2$ on growth, mineral uptake and photosynthesis of NaCl-stressed guava seedlings (*Psidium guajava* L.). *SciHort* 93:125-135.
- [35] Akram MS, M Ashraf and NA Akram. 2009. Effectiveness of potassium sulfate mitigating salt-induced adverse effects on different physio-biochemical attributes in sunflower (*Helianthus annuus* L.). *Flora* 204(6):471-483.
- [36] Azooz MM, MA Shadded and AA Abdel-Latef. 2004. The accumulation and compartmentation of proline in relation to salt tolerance of three sorghum cultivars. *Indian J. Plant Physiol.* 9: 1-8.
- [37] Dagar JC, H Bhagwan and Y Kumar. 2004 Effect on growth performance and biochemical contents of *Salvadora persica* when irrigated with water of different salinity. *Indian J Plant Physiol* 9: 234-238.
- [38] Levit J. 1980. Responses of Plants to Environmental Stresses, Academic Press, Vol. II, New York.
- [39] Jaleel CA, P Manivannan and GMA Lakshmanan. 2007. NaCl as a physiological modulator of proline metabolism and antioxidant potential in *Phyllanthus amarus*. *C R Biologies* 330: 806-813.
- [40] Bota J, J Flexas and H Medrano. 2004. Is photosynthesis limited by decreased Rubisco activity and RuBP content under progressive water stress? *New Phytol.*, 162, pp. 671-681.
- [41] Colebrook EH, SG Thomas, AL Phillips and P Hedden. 2014. The role of gibberellin signalling in plant responses to abiotic stress. *The Journal of Experimental Biology*, 217, 67-75.